

Demonstration of innovative tools



Media & Learning conference - Back to the Future?

Provinciehuis Leuven

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Project Consortium – 5 different partners universities

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Challenges

- Creating more activating videos
- Maximize remote preparation of students and improve learning
- Experimental thinking also for students without lab access
- Encourage active participation by demystifying practical manipulations
- Coping with the continuously increasing student numbers and costs
- Methods to evaluate student progress in this environment







Objectives of the project



To develop educational solutions for students with limited access to laboratory infrastructure

To promote transfer of knowledge using digitally enhanced strategies

To create standardized educational materials

To facilitate educational innovation

To reach out to external stakeholders





What is it all about?

- Virtual lab training for Biomedical science students
- Using an e-learning platform
- Developing a workshop training program
 - Encouraging active thinking and brainstorming
 - Developing a scientific mindset
 - Including self assessment
- Creating
 - A glossary of biomedical terms
 - Student protocols (SOPs)
 - Teacher's guides

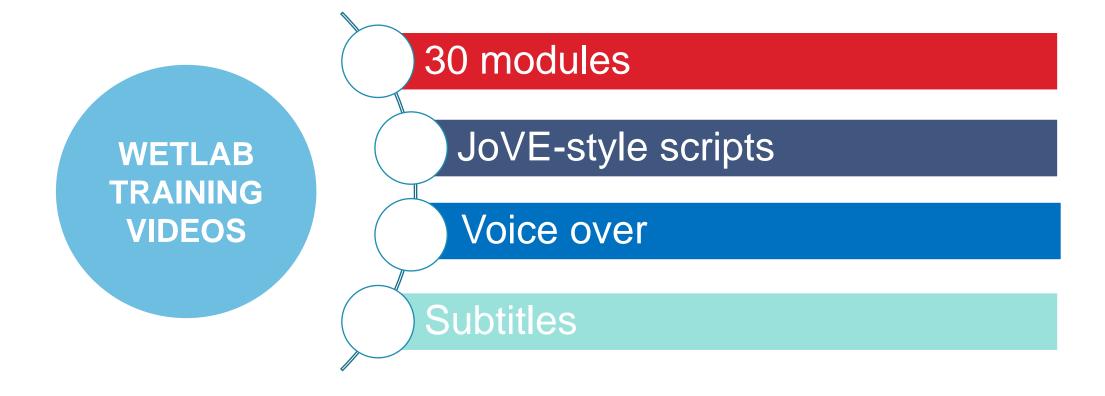
Consortium → Harmonize the quality and format across 5 partners







Virtual lab training for biomedical science students







30 Different modules

KU LEUVEN UIO MUL RU **UNIFG** M1-Chicken embryo staging M1- Zebrafish husbandry & M1- Lypholyzed platelet lysate M1- polarographic methods M1-Zebrafish staging (with developmental staging (with manufacturing for MSC part 1 UiO) M2-Chicken embryo "inking" differentiation MUL) M2- polarographic methods **M2**-Zebrafish injecting Tol2 M3-Chicken embryo "lifting" M2- Zebrafish whole-mount in M2- Osteogenic differentiation part 2 situ hybridization (with KU from MSC and Alizarin red Leuven) **M4-** Mouse gastrulation M3- extracellular flux analysis staining M3-Zebrafish injecting (dissection, staging, EMT techniques part 1 CRISPR-Cas9, Morpholino M3- Toxicological assessment process) M3- Chondrogenic of drugs and compounds M4- extracellular flux analysis differentiation from MSC and M4-7ebrafish small M5-Chicken embryo "beading" techniques part 2 GAG staining compound screen (with M4- Zebrafish behavioral (gain-of-function study) studies (with MUL) UiO) **M5**- mitochondrial morphofunctional analysis by **M6-**Concepts of phenotypic M5-Human tumor cell **M5**-Zebrafish behavioral confocal microscopy analysis of mouse embryos xenografting in larval studies (with UiO) zebrafish M6- preanalytical mass M7- Chorio-allantoic spectrometry membrane (CAM) assay M6-Zebrafish human M6- Zebrafish small cancer cells injection **M8**-Concepts of gene activity M7- mass spectrometry in compound screen (with MUL) metabolomics using reporter mouse lines







30 Different modules

M1-Zebrafish staging (with UiO) M2-Zebrafish injecting Tol2 M3-Zebrafish injecting CRISPR-Cas9, Morpholino M4-Zebrafish small compound screen (with UiO) M5-Zebrafish behavioral studies (with UiO)

M6-Zebrafish human

cancer cells injection

KU LEUVEN UIO

M1-Chicken embryo staging

M2-Chicken embryo "inking"

M3-Chicken embryo "lifting"

M4- Mouse gastrulation (dissection, staging, EMT process)

M5-Chicken embryo "beading" (gain-of-function study)

M6-Concepts of phenotypic analysis of mouse embryos

M7- Chorio-allantoic membrane (CAM) assay

M8-Concepts of gene activity

M1- Ze situ hy Leuvel

M3- To of drug

M4- Ze studies

M5-Hu xenogic zebraf

using reporter mouse lines

M1- Zebrafish husbandry & developmental staging (with M2- Zebrafish whole-mount in situ hybridization (with KU Leuven) **M3**- Toxicological assessment of drugs and compounds M4- Zebrafish behavioral studies (with MUL) M5-Human tumor cell xenografting in larval zebrafish M6- Zebrafish small compound screen (with MUL)

UNIFG RU M1- Lypholyzed platelet lysate M1- polarographic methods manufacturing for MSC part 1 differentiation M2- polarographic methods M2- Osteogenic differentiation part 2 from MSC and Alizarin red M3- extracellular flux analysis staining techniques part 1 M3- Chondrogenic M4- extracellular flux analysis differentiation from MSC and techniques part 2 GAG staining **M5**- mitochondrial morphofunctional analysis by confocal microscopy M6- preanalytical mass spectrometry M7- mass spectrometry in metabolomics







30 Different modules

MUL

- M1-Zebrafish staging (with UiO)
- M2-Zebrafish injecting Tol2
- M3-Zebrafish injecting CRISPR-Cas9, Morpholino
- M4-Zebrafish small compound screen (with UiO)
- M5-Zebrafish behavioral studies (with UiO)
- M6-Zebrafish human cancer cells injection

KU LEUVEN

- M1-Chicken embryo staging
- M2-Chicken embryo "inking"
- M3-Chicken embryo "lifting"
- M4- Mouse gastrulation (dissection, staging, EMT process)
- M5-Chicken embryo "beading" (gain-of-function study)
- M6-Concepts of phenotypic analysis of mouse embryos
- M7- Chorio-allantoic membrane (CAM) assay
- M8-Concepts of gene activity using reporter mouse lines

UiO

- M1- Zebrafish husbandry & developmental staging (with MUL)
- M2- Zebrafish whole-mount in situ hybridization (with KU Leuven)
- M3- Toxicological assessment of drugs and compounds
- M4- Zebrafish behavioral studies (with MUL)
- M5-Human tumor cell xenografting in larval zebrafish
- M6- Zebrafish small compound screen (with MUL)

RU

- M1- Lypholyzed platelet lysate manufacturing for MSC differentiation
- M2- Osteogenic differentiation from MSC and Alizarin red staining
- M3- Chondrogenic differentiation from MSC and GAG staining

UNIFG

- M1- polarographic methods part 1
- **M2** polarographic methods part 2
- M3- extracellular flux analysis techniques part 1
- M4- extracellular flux analysis techniques part 2
- M5- mitochondrial morphofunctional analysis by confocal microscopy
- M6- preanalytical mass spectrometry
- M7- mass spectrometry in metabolomics





PREVIEW

Explantation of a chicken embryo "Lifting"

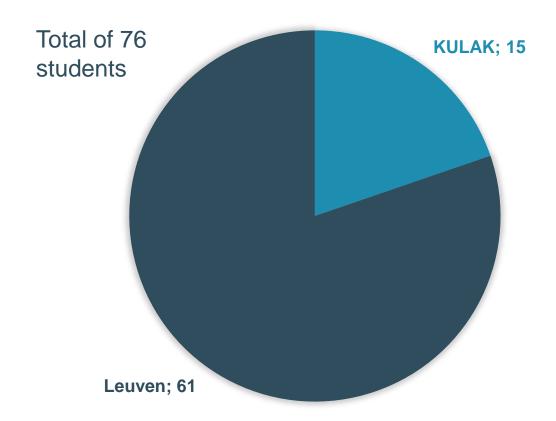


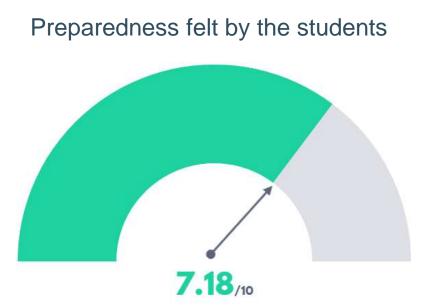






Bèta testers – students 3rd bachelor BMS









Some feedback from the students

"A visual guide is more useful than just written instructions to learn how to perform techniques and manipulations."

"Written instructions are often complex and can be a bit ambiguous, but a demonstration video is clear."

"The added troubleshooting is also useful especially when these problems occur often in the practical course."

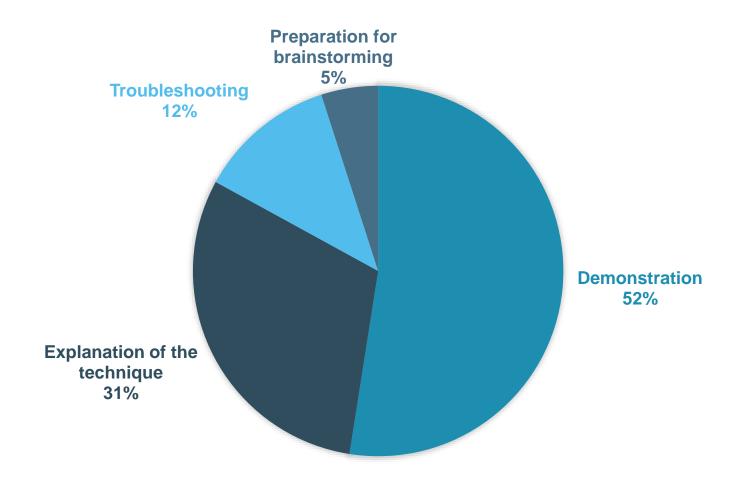
"I felt more prepared and more capable going into the practical course."







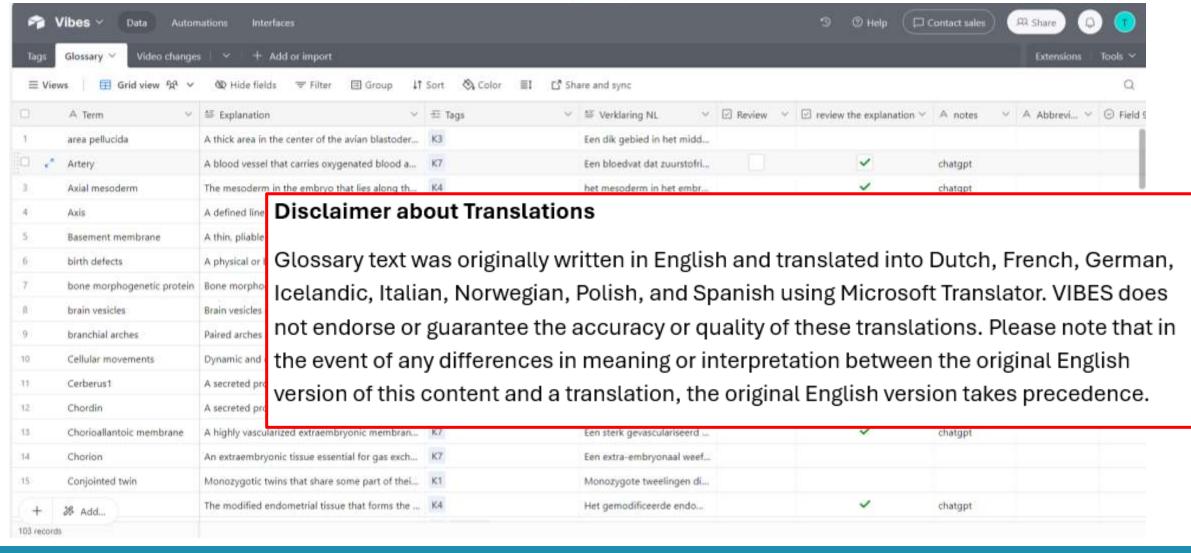
What exactly in the video was helpful?





Glossary







Student protocols (SOP)

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Disclaimer:

The educational material provided herein is intended solely for teaching purposes, in the context of higher education in biomedical science and related disciplines. Any hands-on activities related to this material should be done only under the supervision of qualified educational staff. It is the responsibility of the user to perform experiments in an appropriate laboratory context and to take into account local safety, health, and environmental legislation.







Teacher's guide

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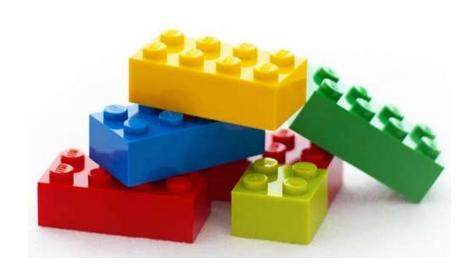
How to share materials?

VIBES provide all the lego parts on their website

→ Accessible for teachers and students only

H5P?

- Incompatible
- Licence issues



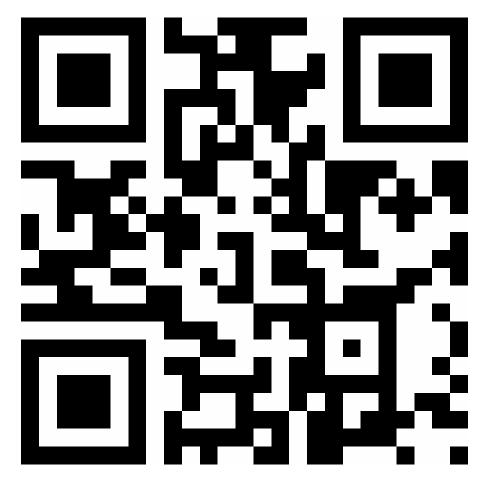
Solution: interactive powerpoint used by all partner universities on their own learning platforms







More info and conference data & venue



Home (vibes-edu.eu)







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